Development of imaging approaches to monitor tumor-reactive T cells in vivo

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Kristine Mayer



Dario Gosmann

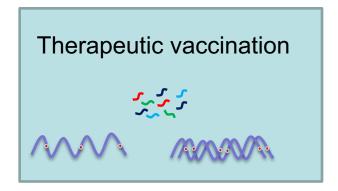


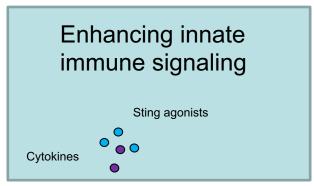
Lisa Russelli

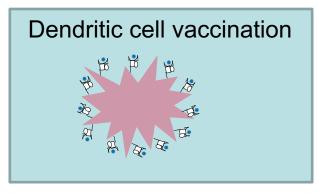


Theresa Käsbauer Sandro Bissenberger

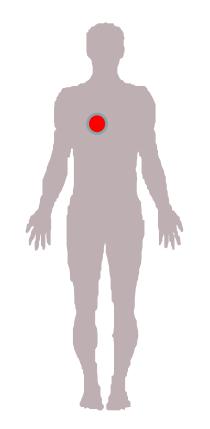
Cancer Immunotherapy

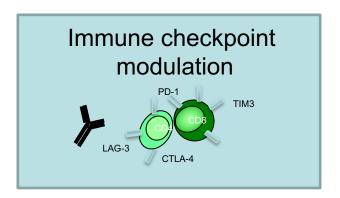


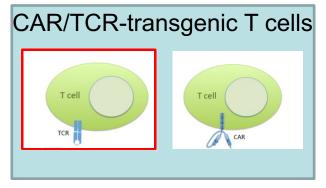


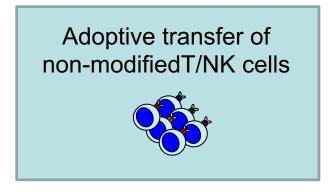


Patient with cancer

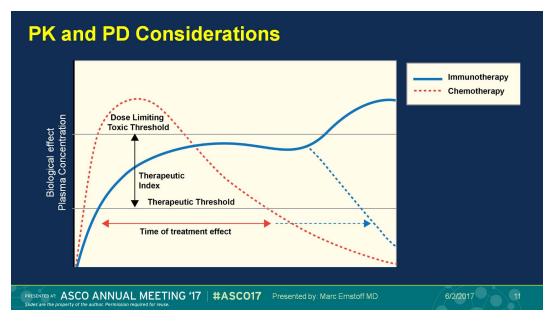


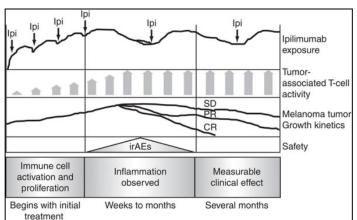


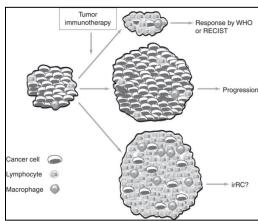




Unique characteristics of Immunotherapy







G. Pennock, et.al, American Journal of Clinical Oncology. 2012

Research project C10

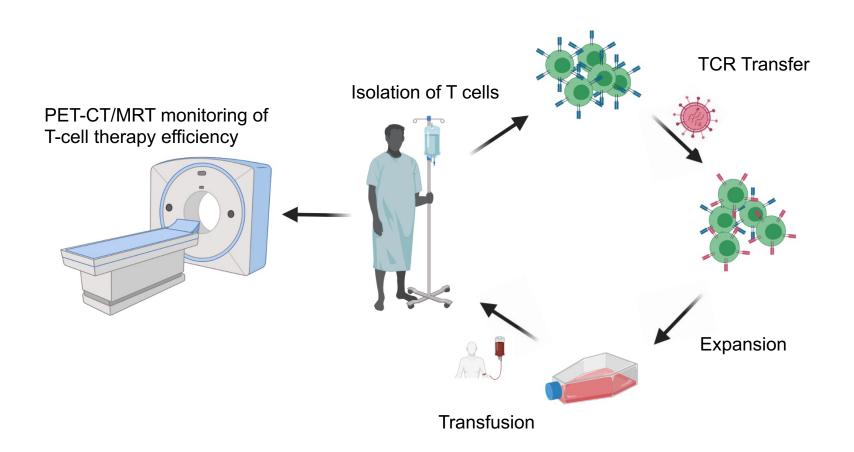
Development of imaging techniques to monitor tumor reactive T cells in vivo based on antibody-derived constructs

Tracer development

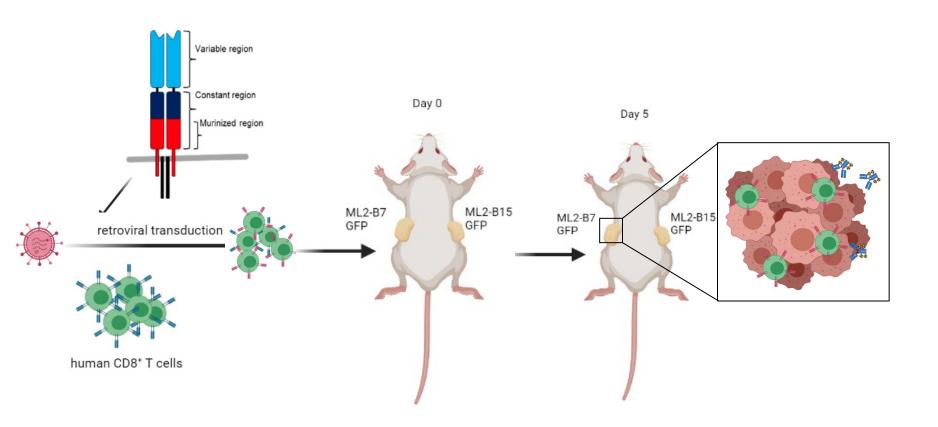
Metabolic imaging of T cell activity in tumors

BLI imaging of T cells

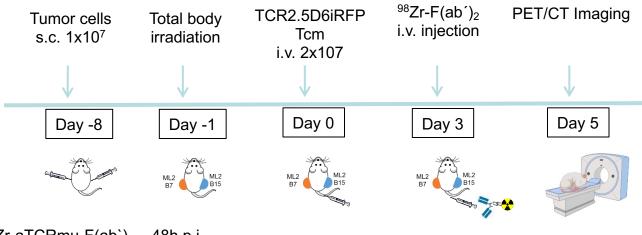
Adoptive T-cell transfer of TCR-transgenic T cells



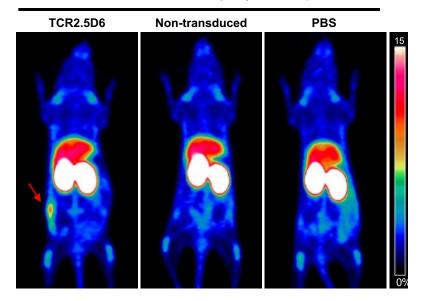
Principle of Immuno-Imaging of TCR-transgenic T cells

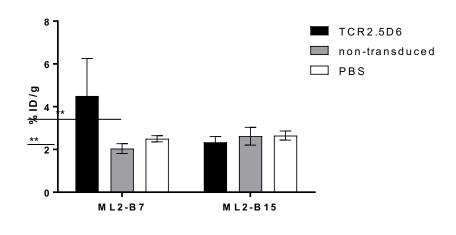


In vivo imaging of T cells by 89Zr-aTCRmu-F(ab`)₂ by Immuno-PET



 89 Zr-aTCRmu-F(ab`)₂ – 48h p.i.



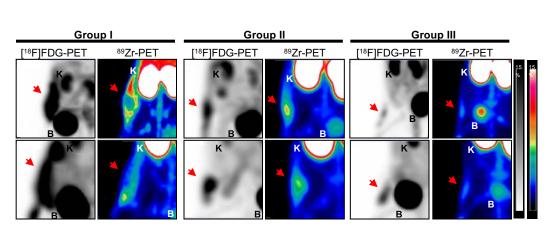


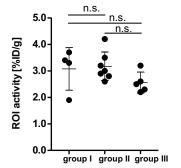
Mall et al. Cancer Research 2016 Yusufi et al. Theranostics 2017

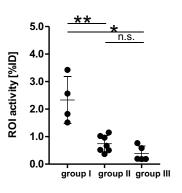
Identification of heterogeneity of T-cell infiltration by Immuno-PET

Tumor cells s.c. 1x107

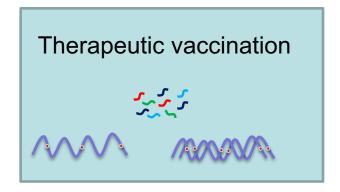
Group I Group III							
D-10	D-8	D-6	D-1	D+0 D+1	D+3	D+3	D+5
			TBI 1Gy	T _{CM} 1.5x10 ⁷ i.v.	¹⁸ F-FDG Imaging	⁸⁹ Zr-aTCRmu-F(ab`) ₂ i.v.	PET/CT Imaging

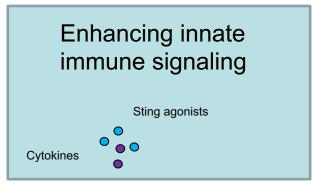


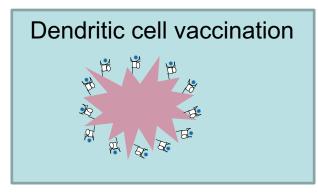




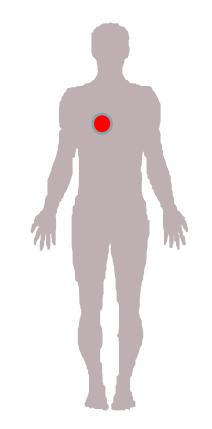
Cancer Immunotherapy

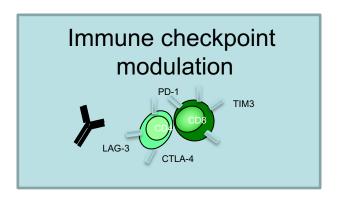


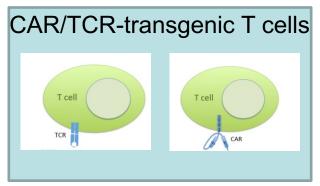


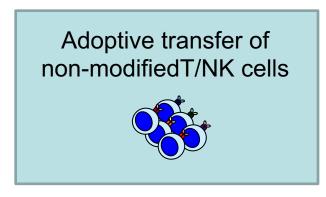


Patient with cancer

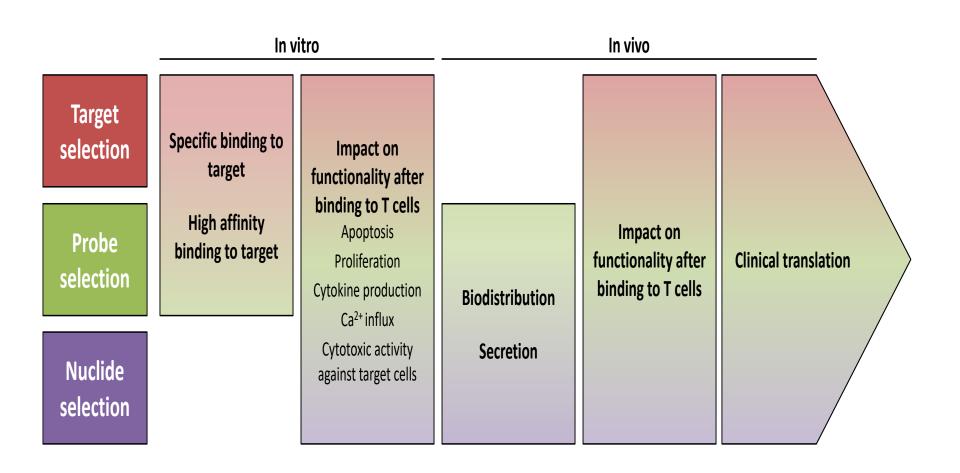






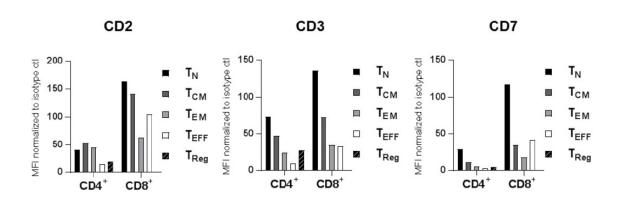


General T-Cell monitoring by Immuno-PET

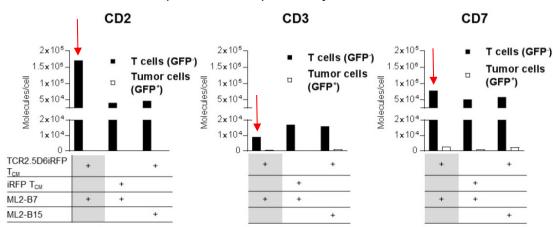


Target selection

Selection of suitable surface markers

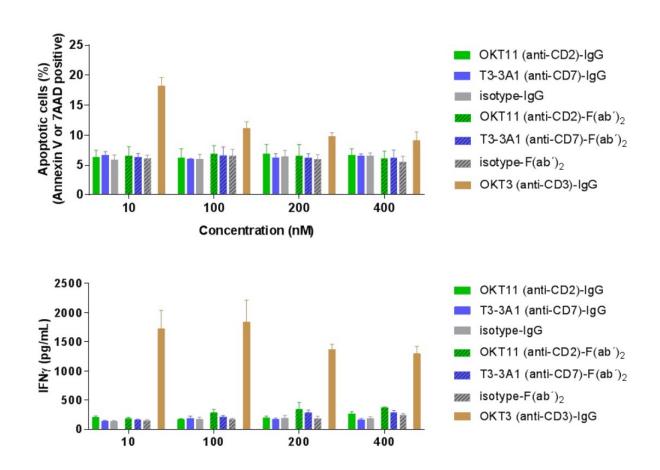


Expression on specifically activated T-cells



Target selection

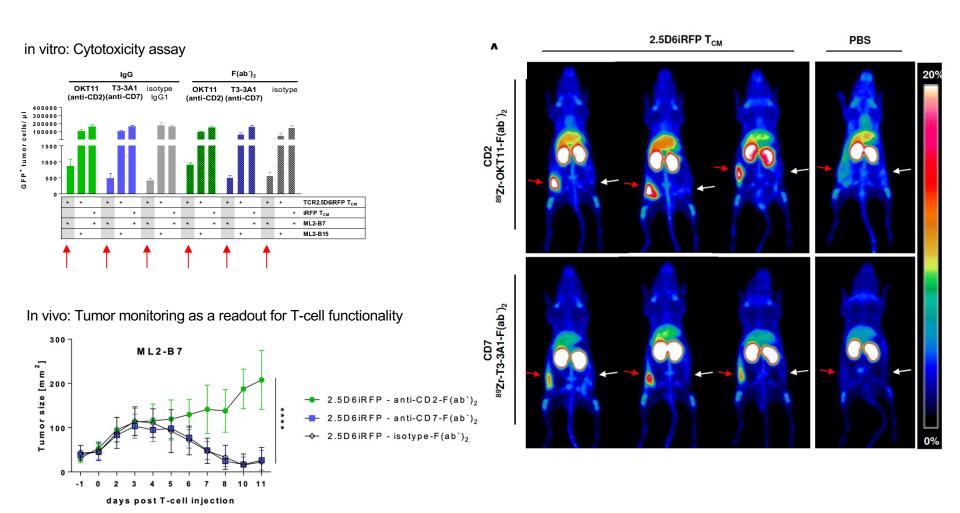
Impact of selected antibody-derived constructs on T cell function in vitro





Target selection

Impact of selected antibody-derived constructs on T cell function in vitro and in vivo





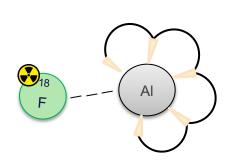


Nuclide selection

Development of novel tracers: Al¹⁸F method

Radioisotope of choice → F-18

- Maximum positron energy of 0.635 MeV → high resolution PET images
- Half-life of 109.8 min → avoid prolonged irradiation of subjects but allow in vivo investigations

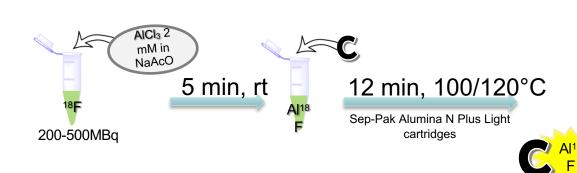


Radiofluorination method → AIF-18

Al forms octahedral complexes → pentadentate ligand is desired with negatively charged oxygen donor groups, leaving only one binding site open for the F-18 ion

NOTA

- Radiolabeling T ~ 100°
- RCY ≥ 75%
- Serum stability (after 4h) → 87%
- High in vivo stability



Nuclide selection

AMP-based chelator Al¹⁸Fmethod

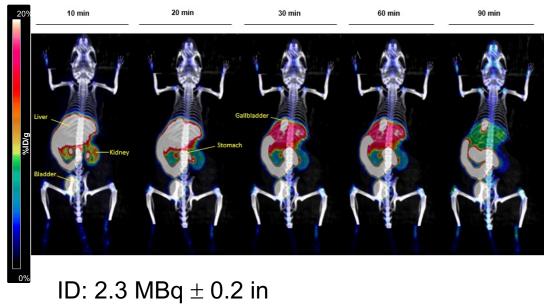
- Radiochemical yield of 55% at pH 5 and room temperature.
 High stability in Human
- Serum was measured for [Al¹⁸F(2-AMPDA-HB)]⁻, with 90±4% of F-18 complexed after 2h and 87±5% after 4h.



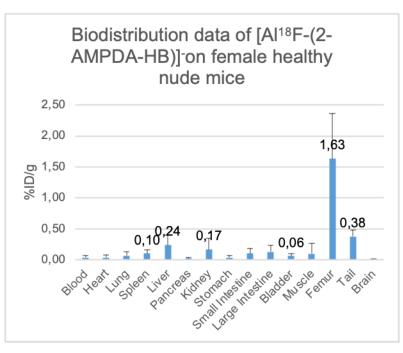




In vivo PET/CT dynamic scan



ID: 2.3 MBq ± 0.2 in 0.9% NaCl



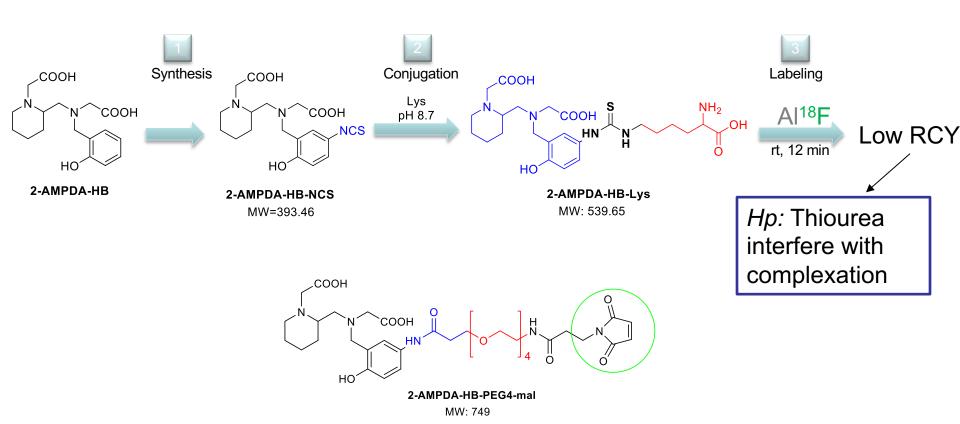
Mice sacrificed 2h p.i.



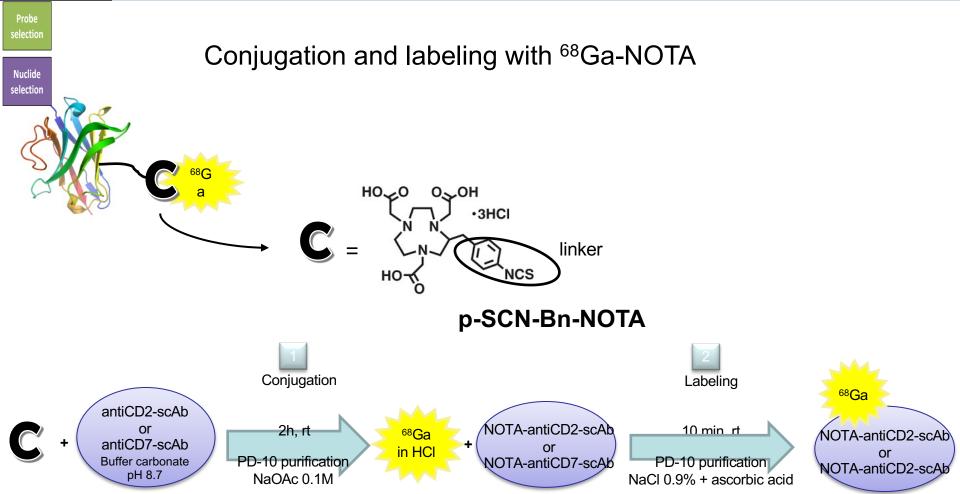
Probe selection

Nuclide selection

Functionalised 2-AMPDA-HB







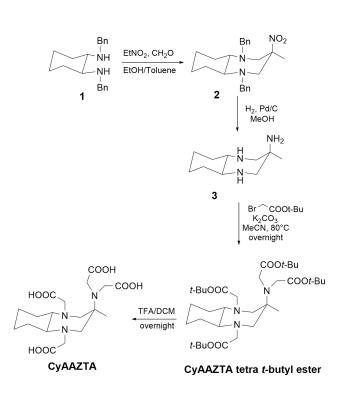


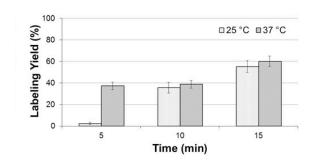




CyAAZTA publication Tei/D`Alessandria

"A rigidified AAZTA-like ligand as efficient chelator for 68Ga radiopharmaceuticals"





RCY at rt, 15min \rightarrow 80.7% RCY at 90°C, 5min \rightarrow 94.7%

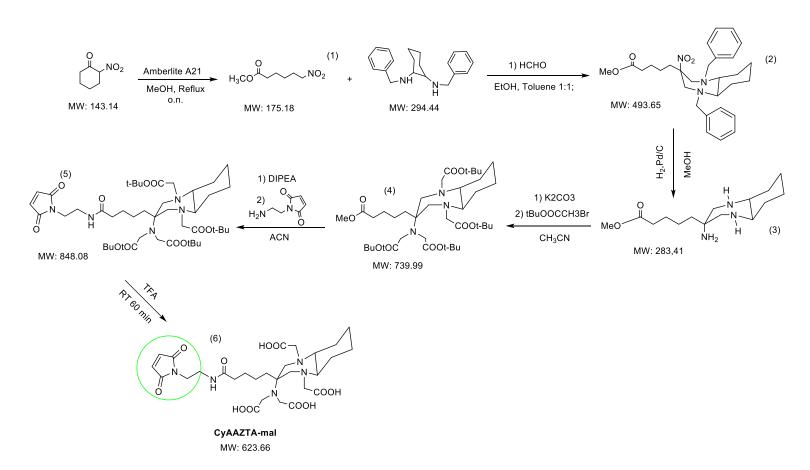
⁶⁸GaCyAAZTA is stable in HS and DTPA over 90 minutes (HPLC data)



Probe selection

Nuclide selection

CyAAZTA functionalized synthesis

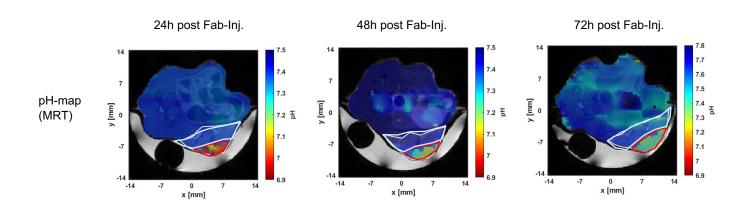


Metabolic pH imaging

Imaging of pH in vivo using hyperpolarized ¹³C-labelled cymonic acid

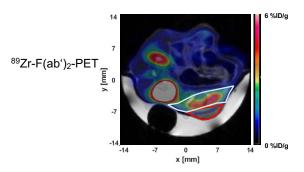


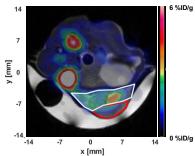
Franz Schilling

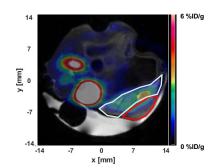




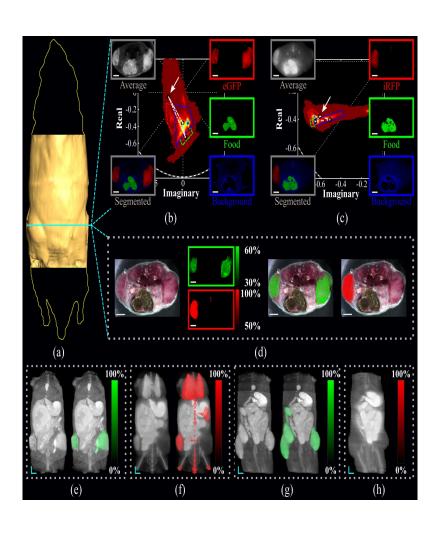
Martin Grashei







Multispectral epi-illumination cryoslicing fluorescence imaging



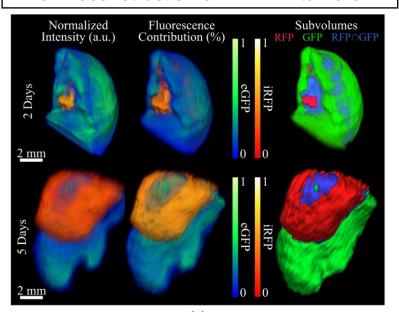




Vasili Ntziachristos

Dimitri Gkorpas

3D reconstruction of ML2-B7 tumors



Publications

Robu S, Richter A, Gosmann D, Seidl C, Leung D, Hayes W, Cohen D, Morin P, Donnelly DJ, Lipovšek D, Bonacorsi SJ, Smith A, Steiger K, Aulehner C, **Krackhardt AM**, Weber WA: Synthesis and Preclinical evaluation of 68Ga-labeled Adnectin, 68Ga-BMS-986192 as a PET agent for Imaging PD-L1 expression. J Nucl Med 2021, doi: 10.2967/jnumed.120.258384

Russelli L, Martinelli J, De Rose F, Reder S, Herz M, Schwaiger M, Weber W, Tei L, **D'Alessandria C**. Room Temperature Al18F Labeling of 2-Aminomethylpiperidine-Based Chelators for PET Imaging. ChemMedChem. 2020 Feb 5;15(3):284-292.

Audehm S, Glaser M, Pecoraro M, Bräunlein E, Mall S, Klar R, Effenberger M, Albers J, Bianchi HO, Peper J, Yusufi N, Busch DH, Stevanovic S, Mann M, Antes I, **Krackhardt AM**: Key features relevant to select antigens and TCR from the MHC mismatched repertoire to treat cancer; Front Immunol 2019; 10: 1485; doi: 10.3389/fimmu.2019.01485

Albers J, Ammon T, Gosmann D, Audehm A, Thoene S, Winter C, Secci R, Wolf A, Stelzl A, Steiger K, Ruland J, Bassermann F, Kupatt C, Anton M, and **Krackhardt AM**: Gene editing enables T cell engineering to redirect antigen specificity for potent tumor rejection; Life Science Alliance 2019, doi: 10.26508/lsa.201900367.

Mayer KE, Mall S, Yusufi N, Gosmann D, Steiger K, Russelli L, Bianchi H, Audehm S, Wagner R, Bräunlein E, Stelzl A, Bassermann F, Weichert W, Weber W, Schwaiger M, D`Alessandria C, **Krackhardt AM**: T-cell functionality testing is highly relevant to developing novel immuno-tracers monitoring T cells in the context of immunotherapies and revealed CD7 as an attractive target; Theranostics 2018; 8: 6070-6087.

Varasteh Z, De Rose F, Mohanta S, Li Y, Zhang X, Miritsch B, Scafetta G, Yin C, Sager HB, Glasl S, Gorpas D, Habenicht AJR, Ntziachristos V, Weber WA, Bartolazzi A, Schwaiger M, **D'Alessandria C**. Imaging atherosclerotic plaques by targeting Galectin-3 and activated macrophages using (89Zr)-DFO-Galectin3-F(ab')2 mAb. Theranostics. 2021 Jan 1;11(4):1864-1876.

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Sybille Reder

Markus Mitterlhäuser

Hannes Rolbieski

Michael Herz

Biomedical Magnetresonance

Franz Schilling

Martin Grashei

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Lorenzo Tei

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